

**SYNTHESIS OF A POTENT ANTITHROMBIN ACTIVATING PENTASACCHARIDE:  
A NEW HEPARIN-LIKE FRAGMENT CONTAINING TWO 3-O-SULPHATED GLUCOSAMINES**

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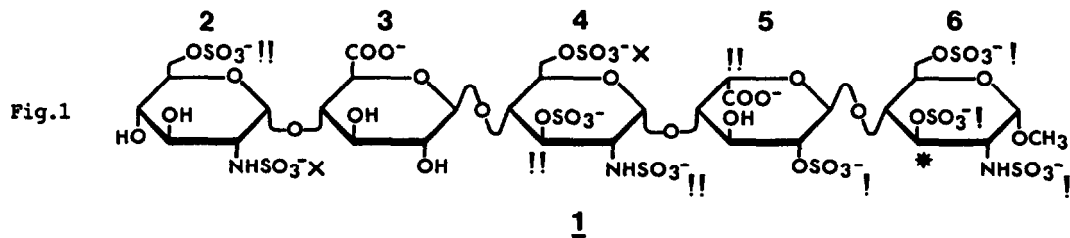
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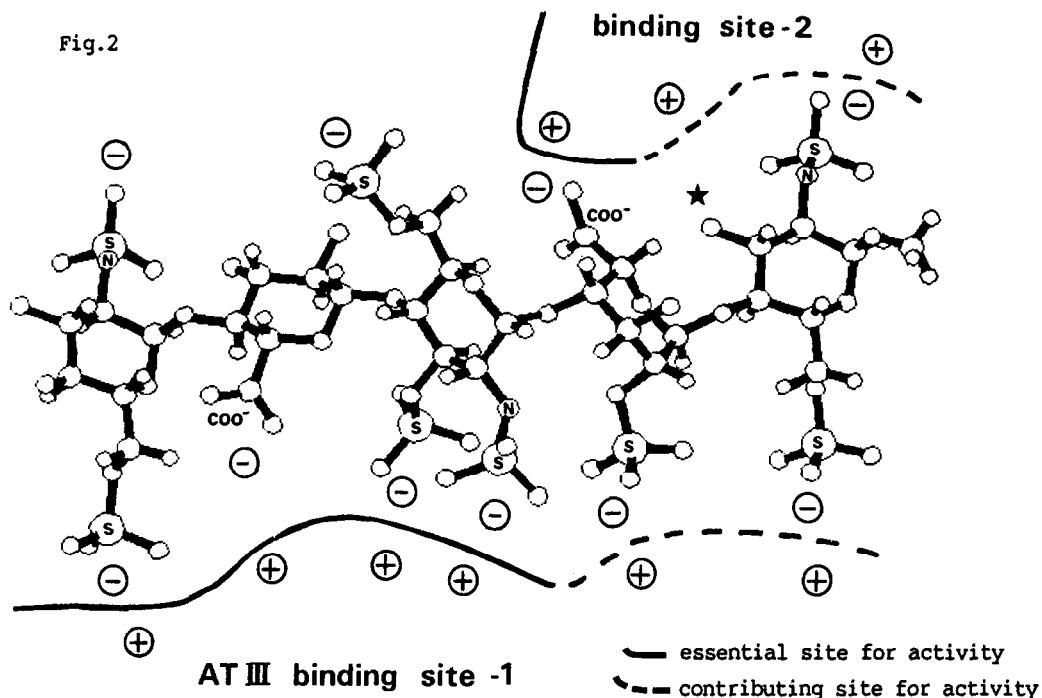
**Summary**

The synthesis of a pentasaccharide corresponding to the antithrombin III binding region of heparin, but containing an extra 3-O-sulphate group at the reducing end, is described. This compound elicits higher anti-Xa activity than the antithrombin III binding region of heparin.

It is well established now that the minimal antithrombin III (AT-III) binding region of heparin consists of an unique pentasaccharide fragment<sup>1</sup>. This pentasaccharide, which has become synthetically available<sup>2</sup>, catalyzes the AT-III mediated inactivation of factor Xa (anti-Xa activity), but not of thrombin.

In this communication we wish to introduce a very potent synthetic analogue (i.e. compound 1 in Fig. 1) of the naturally occurring fragment, containing an additional 3-O-sulphate group at the reducing glucosamine-unit 6<sup>3</sup> (see asterisk in Fig.1). This analogue displays an anti-Xa activity of about 1270 U/mg in an amidolytic assay<sup>4</sup>, whereas the synthetic pentasaccharide corresponding to the AT-III binding site of heparin, displays 590 anti-Xa U/mg<sup>5</sup>. The higher activity of analogue 1 has to be attributed to the presence of the additional 3-O-sulphate group. In this respect, it is important to note that the 6-O-sulphate group at glucosamine unit 2<sup>6</sup>, the 3-O-sulphate<sup>7</sup> and N-sulphate<sup>8</sup> groups at glucosamine unit 4, as well as the carboxylate moiety of iduronic acid unit 5<sup>9</sup> are essential (!! in Fig. 1) for activation of AT-III. In addition, the 2-O-sulphate at iduronic acid unit 5<sup>10</sup> and the 6-O-sulphate<sup>11</sup> and N-sulphate<sup>8</sup> groups of glucosamine unit 6 increase (! in Fig. 1) the AT-III mediated activity. On the other hand the N-sulphate (N-acetyl) group at unit 2<sup>1b,7</sup> and the 6-O-sulphate at unit 4<sup>12</sup> are considered to be non-essential (x in Fig. 1).



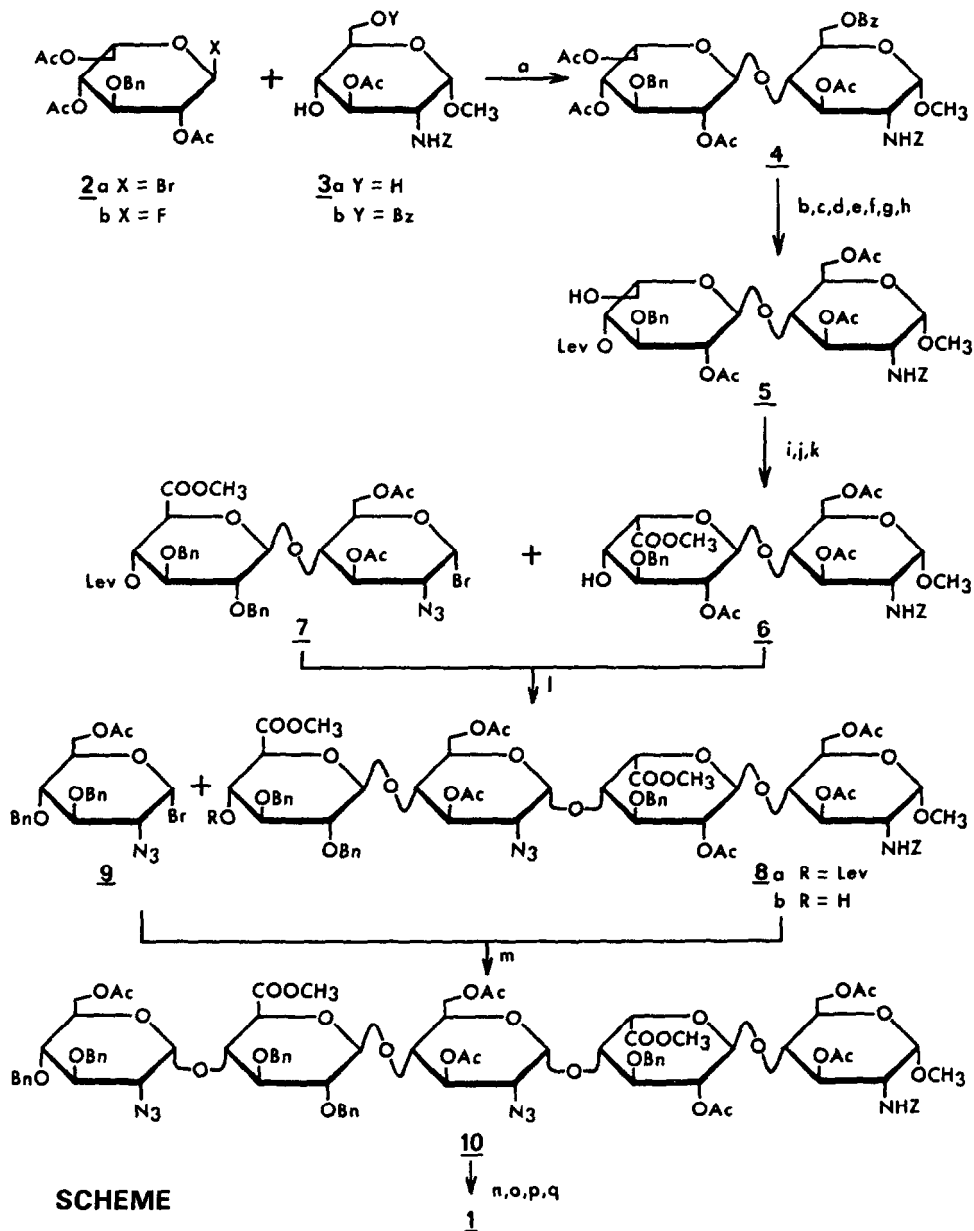


Taking into consideration these findings, the heparin-binding site of AT-III can be mapped around a molecular model<sup>9a,13</sup> of the naturally occurring heparin pentasaccharide (Fig. 2). In Fig.2 one can see that the pentasaccharide exhibits a linear conformation with binding areas at the south- and north site of the molecule (i.e. AT-III binding sites 1 and 2, respectively). The higher activity of compound 1, relative to the naturally occurring heparin fragment, can be tentatively ascribed to its enhanced interaction with AT-III at binding site-2, brought about by the extra 3-O-sulphate group (In Fig.2 this sulphate group would be located at the position of the asterisk  $\star$ ).

The synthesis of compound 1 is outlined in the Scheme. Following a well-established strategy<sup>2,9,11</sup> the fully protected pentasaccharide 10 should be prepared, containing acetyl functions at hydroxyl functions to be sulphated and benzyl protective groups for unsulphated hydroxyl groups.

We started the synthesis<sup>14</sup> from easily available methyl 4,6-O-benzylidene-2-benzoyloxycarbonyl-amino-2-deoxy- $\alpha$ -D-glucopyranoside<sup>15</sup>, which was acetylated in acetic anhydride/pyridine and then treated with aqueous acetic acid to afford 3a (90% yield). Compound 3a was selectively benzoylated at the primary hydroxyl group to afford compound 3b in 79% yield.

Since coupling of the unreactive compound 3b with known L-idopyranosyl bromide derivative 2a<sup>2b</sup> was disappointing we turned attention to the corresponding fluoride derivative 2b. Treatment of 1,2,4,6-tetra-O-acetyl-3-O-benzyl-L-idopyranose<sup>2b</sup> with 70% hydrogen fluoride/pyridine in dichloromethane for 4 h at 0°C, gave after work-up and chromatography pure 2b in 60% yield (<sup>1</sup>H-NMR (CDCl<sub>3</sub>),  $\delta$  = 5.70 (dd, J=48 Hz, J=3Hz, H-1). Condensation of compounds 2b and 3b in the presence of borontrifluoride etherate gave disaccharide 4. Conversion of 4 into the desired iduronic acid-glucosamine building block 6 has been performed as described recently in similar syntheses<sup>2b,11</sup>. First, compound 4 was re-protected in seven steps to give 5. Oxidation of compound 5 with chromium (vi) oxide was followed by diazomethane treatment and deprotection of the levulinoyl ester to give compound 6 ( $[\alpha]_D^{20}$  = 20.5, c = 1, dichloromethane).



Scheme: a)  $\text{BF}_3$ -etherate, MS 4A,  $\text{CH}_2\text{Cl}_2$ ,  $-20^\circ\text{C}$ , (73%); b)  $\text{KOTBu}$ ,  $\text{MeOH}/\text{dioxane}$ , RT; c) 2,2-dimethoxy propane,  $\text{pTS}$ ,  $\text{DMF}$ , RT, (b $\rightarrow$ c, 90%); d)  $\text{Ac}_2\text{O}$ , pyridine,  $35^\circ\text{C}$ ; e)  $\text{AcOH}/\text{H}_2\text{O}$ ,  $40^\circ\text{C}$ , (d $\rightarrow$ e, 93%); f) Dimethoxytrityl chloride,  $\text{THF}/\text{pyridine}$ ,  $-6^\circ\text{C}$ ; g) Levulinic acid anhydride,  $\text{THF}/\text{pyridine}$ , RT; h)  $\text{AcOH}/\text{H}_2\text{O}$ , RT, (f $\rightarrow$ h, 85%); i)  $\text{CrO}_3$ , acetone,  $0^\circ\text{C}$ ; j)  $\text{CH}_2\text{N}_2$ ,  $\text{CH}_2\text{Cl}_2$ , RT, (i $\rightarrow$ j, 85%); k)  $\text{H}_2\text{NNH}_2$ ,  $\text{AcOH}$ , pyridine, RT, (83%); l)  $\text{AgSO}_3\text{CF}_3$ , MS 10A,  $\text{CH}_2\text{Cl}_2$ ,  $-30^\circ\text{C}$ , (50%); m)  $\text{AgSO}_3\text{CF}_3$ , MS 4A, 2,6-di-*t*-butylpyridine,  $\text{CH}_2\text{Cl}_2$ ,  $-55^\circ\text{C}$ , (80%); n)  $\text{NaOH}$ ,  $\text{H}_2\text{O}/\text{MeOH}/\text{CHCl}_3$ , RT, (55%); o)  $\text{SO}_3\text{N}(\text{CH}_3)_3$ ,  $\text{DMF}$ ,  $50^\circ\text{C}$ , (55%); p)  $\text{H}_2$ ,  $\text{Pd}/\text{C}$ ,  $\text{MeOH}/\text{H}_2\text{O}$ , RT; q)  $\text{SO}_3\text{N}(\text{CH}_3)_3$ ,  $\text{Na}_2\text{CO}_3$ ,  $\text{H}_2\text{O}$ , RT, (p $\rightarrow$ q, 70%).

Abbreviations: Bn = benzyl, Bz = benzoyl, Ac = acetyl, Lev = levulinoyl, Z = benzyloxycarbonyl

Reaction of **6** with known glycon **7**<sup>2b</sup>, in the presence of silver triflate and molecular sieves, gave tetrasaccharide **8a**. Compound **8b** was obtained after hydrazinolysis of the levulinoyl group. Coupling of excess of **9** with **8b** in the presence of silver triflate and 2,6-di-*t*-butylpyridine gave, after purification, the fully protected pentasaccharide **10** ( $[\alpha]_D^{20} = 44.7$ ;  $c = 0.9$ , dichloromethane)<sup>16</sup>. Finally, in the following four-step procedure the fully protected derivative **10** was converted into analogue **1**: i) simultaneous saponification of acetyl esters and carboxyl-methyl esters; ii) O-sulphation, Sephadex LH-20 chromatography; iii) hydrogenolysis; iv) N-sulphation. The crude product was purified by Sephadex DEAE chromatography and then desalted (Sephadex G10). The structure of compound **1** was confirmed by 2-dimensional proton-proton correlated spectroscopy (2D-COSY)<sup>17</sup>. Most remarkably, the  $\alpha$ -L-iduronic acid part of compound **1** adopts (in D<sub>2</sub>O) mainly the <sup>2</sup>S<sub>0</sub> skew boat conformation<sup>13b,18</sup>, whereas  $\alpha$ -L-iduronic acid in the natural occurring pentasaccharide occurs in an equilibrium between <sup>2</sup>S<sub>0</sub> and <sup>1</sup>C<sub>4</sub> forms (ratio about 2:1)<sup>19,20</sup>.

#### Acknowledgement

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3. The unique pentasaccharide is part of an isolated octasaccharide (ref. 1) 1-[2-3-4-5-6]-7-8:-Ida-[GlcNAc(6-OSO<sub>3</sub>)-Glc-GlcNSO<sub>3</sub>(3,6-OSO<sub>3</sub>)-Ida(2-0-SO<sub>3</sub>)-GlcNSO<sub>3</sub>(6-O-SO<sub>3</sub>)]-Ida(2-OSO<sub>3</sub>)-A Man.
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5. The synthetic AT-III binding site of heparin, containing  $\alpha$ -methoxy at the anomeric center, elicits 740 anti-Xa U/mg (Th.G. van Dinther, unpublished result).
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- b. Although 2-O-sulphated  $\alpha$ -L-iduronic acid in heparin-like fragments occur in an equilibrium between <sup>2</sup>S<sub>0</sub> and <sup>1</sup>C<sub>4</sub> forms, we postulated that the pentasaccharide bound at AT-III displays  $\alpha$ -L-iduronic acid in the <sup>1</sup>C<sub>4</sub> conformation (see ref. 20).
14. Currently we introduce a methyl group at the anomeric center to avoid side-reactions at the reducing end during the last two steps of the synthesis (refs. 9 and 11).
15. S. Akiya and T. Osawa, *Yakugaku Zasshi*, **6**, 1276 (1956).
16. <sup>1</sup>H-NMR (360 MHz, CDCl<sub>3</sub>), compound **10**,  $\delta = 4.68$  (d, J = 3.5 Hz, H-1), 5.06 (d, J = 2.5 Hz, H-1'), 4.92 (d, J = 3.5 Hz, H-1''), 4.35 (d, J = 7.9 Hz, H-1'''), 5.50 (d, J = 3.6 Hz, H-1'''').
17. <sup>1</sup>H-NMR (360 MHz, D<sub>2</sub>O) compound **1**,  $\delta = 4.97$  (d, J = 3.5 Hz, H-1); 5.10 (d, J = 5.3 Hz, H-1'); 5.58 (d, J = 3.5 Hz, H-1''); 4.59 (d, J = 7.9 Hz, H-1'''); 5.59 (d, J = 3.5 Hz, H-1'''''); 4.38 (dd, J = 10.8 Hz, J = 9.2 Hz, H-3); 4.32 (dd, J = 10.8 Hz, J = 9.2 Hz, H-3'').  $[\alpha]_D^{20} = 38.4$  ( $c = 0.61$ , H<sub>2</sub>O).
18. The  $\alpha$ -L-iduronic acid part of the spectrum was computer-simulated (PANIC.84): J 1,2 = 5.3 Hz, J 2,3 = 9.4 Hz, J 3,4 = 4.4 Hz, J 4,5 = 4.4 Hz (<sup>2</sup>S<sub>0</sub> : <sup>1</sup>C<sub>4</sub> is about 9:1).
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